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# STUDIES ON INHIBITORS OF RAT MAST CELL DEGRANULATION PRODUCED BY MICROORGANISMS

### II. STRUCTURE ELUCIDATION OF EUROCIDINS D AND E

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The planar structures of new eurocidin related compounds, eurocidins D and E, were elucidated from  ${}^{1}H{}^{-1}H$  shift correlated 2D NMR spectra and other NMR data. All protons in the molecules were assigned. Eurocidins D and E have novel pentaenic structures of eurocidin family.

In the preceding paper<sup>1</sup>), screening of microorganisms, isolation and physico-chemical properties of new eurocidin related compounds, eurocidins C, D and E have been described. These compounds were isolated from the culture broth of *Streptoverticillium eurocidicum* IFO 13491 as the potent inhibitors of mast cell degranulation induced by compound 48/80.

We report here the structure elucidation of eurocidins D and E, the major components of them, from the NMR data and other physico-chemical properties.

Fig. 1. Structures of eurocidins D and E.



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## **Results and Discussions**

## <sup>13</sup>C NMR Spectra

The <sup>13</sup>C NMR data of eurocidins D and E are shown in Table 1. The measurement of multiplicity was carried out with DEPT experiments ( $\theta$ =45°, 90° and 135°). It was estimated from elemental analysis and the mass spectral data in the preceding paper<sup>1</sup>) that the carbon number of eurocidins D and E is 40. The signals of 38 and 39 carbons in eurocidins D and E, respectively, were observed in their <sup>13</sup>C NMR spectra because of mutual superposition in chemical shifts of conjugated double bond carbons at

$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	Carbon	Chemical shift (ppm), multiplicity				
Ketone         208.7 s         208.0 s           COOH         176.8 s         176.8 s         176.1         177.6           Lactone         169.9 s         169.9 s         170.3         170.1           CH         133.6 d         133.6 d         133.6 d         170.3         170.1           CH         133.7 d         133.6 d         133.6 d         170.3         170.1           CH         133.7 d         133.6 d         133.6 d         170.3         170.1           CH         133.7 d         133.6 d         133.5 d         170.3         170.1           CH         133.7 d         133.6 d         170.3         170.1         170.1           CH         132.5 d         133.5 d         170.3         170.1         170.1         170.1           CH         132.5 d         133.5 d         170.3         170.1		Eurocidin D	Eurocidin E	Amphotericin A <sup>a</sup>	Amphotericin B	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Ketone	208.7 s	208.0 s			
Lactone $169.9 \text{ s}$ $170.3$ $170.1$ -CH $136.6 \text{ d}$ $136.6 \text{ d}$ $170.3$ $170.1$ -CH $133.7 \text{ d}$ $133.5 \text{ d}$ $133.5 \text{ d}$ $133.5 \text{ d}$ -CH $133.5 \text{ d}$ $133.5 \text{ d}$ $133.5 \text{ d}$ $133.5 \text{ d}$ -CH $d$ $d$ $d$ $d$ -CH $131.3 \text{ d}$ $131.4 \text{ d}$ $129 \sim 136$ -CH $130.9 \text{ d}$ $131.1 \text{ d}$ $129 \times 136$ -CH $129.8 \text{ d}$ $1228.8 \text{ d}$ $97.0$ $97.1$ Acetal $97.0 \text{ s}$ $97.0$ $97.1$ $97.9$ $97.9$ $97.9$ CH $72.7 \text{ d}$ $72.6 \text{ d}$ $72.6 \text{ d}$ $55.9 \text{ d}$ $55.9 \text{ d}$ $55.9 \text{ d}$ $55.9 \text{ d}$ $55.60 \text{ d}$ $66 \sim 76 \text{ d}$ $66 \sim 76 \text{ d}$ $66 \times 38 \times 39 \text{ d}$ $66 \times 38 \times 39 \text{ d}$ $72.9 \text{ d}$ </td <td>COOH</td> <td>176.8 s</td> <td>176.8 s</td> <td>176.1</td> <td>177.6</td>	COOH	176.8 s	176.8 s	176.1	177.6	
$ \begin{array}{c} -CH & 136.6 d & 136.6 d \\ = CH & 133.7 d & 133.6 d \\ = CH & 133.5 d & 133.5 d \\ = CH & d & d \\ = CH & 131.3 d & 131.4 d \\ = CH & 130.9 d & 131.1 d \\ = CH & 129.8 d & 129.8 d \\ = CH & 129.8 d & 129.8 d \\ = CH & 128.6 d & 128.5 d \\ = CH & 128.6 d & 128.5 d \\ = CH & 128.6 d & 128.5 d \\ = CH & 128.6 d & 128.5 d \\ = CH & 128.6 d & 128.5 d \\ = CH & 128.6 d & 128.5 d \\ = CH & 75.0 d & 74.9 d \\ CH & 74.2 d & 74.1 d \\ CH & 72.7 d & 72.6 d \\ CH & 70.7 d & 69.9 d \\ CH & 70.7 d & 69.9 d \\ CH & 65.6 d & 65.5 d \\ CH & 66.6 d & 56.1 d \\ CH & 38.3 d & 58.2 d \\ CH & 38.1 d & 58.2 d \\ CH & 38.5 d & 38.4 d \\ CH_2 & 44.7 t & 46.0 t \\ CH_2 & 43.5 t & 44.6 t \\ CH_2 & 36.6 t & 37.9 t \\ CH_2 & 26.3 t & 35.1 t \\ CH_2 & 25.3 t & 25.2 t \\ CH_3 & 17.9 q & 17.9 q \\ CH_3 & 17.9 q & 17.9 q \\ CH_4 & 14.4 q \\ 114.q \\ CH_4 & 114.q \\ CH_4 & 114.q \\ \end{array}$	Lactone	169.9 s	169.9 s	170.3	170.1	
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	=CH	d	131.2 d			
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CH       75.0 d       74.9 d         CH       74.2 d       74.1 d         CH       72.7 d       72.6 d         CH       70.7 d       69.9 d         CH       70.0 d       67.9 d         CH       66~76         CH       65.6 d       65.7 d         CH       65.5 d       65.5 d         CH       65.5 d       65.5 d         CH       66~76         CH       65.6 d       38~39         CH       65.5 d       65.5 d         CH       65.1 d       56.1 d         CH       56.1 d       56.1 d         CH2       48.9 t       49.0 t         CH2       43.5 t       44.6 t         CH2       43.5 t       44.6 t         CH2       43.5 t       38.4 d         CH2       43.5 t       40~45         CH2       43.5 t       40~45         CH2       35.1 t       36.6 t         CH2       25.3 t       25.2 t         CH2       25.3 t	Acetal	95.9 d	95.8 d	97.3	95.9	
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CH       72.5 d         CH       70.7 d       69.9 d         CH       70.0 d       67.9 d         CH       68.0 d       67.5 d         CH       65.6 d       66~76         CH       65.6 d       65.6 d         CH       65.5 d       65.5 d         CH       64.1 d       63.9 d         CH       56.1 d       58.2 d         CH       56.1 d       56.1 d         CH2       48.9 t       49.0 t         CH2       48.9 t       49.0 t         CH2       44.7 t       46.0 t         CH2       43.5 t       44.6 t         CH2       43.5 t       44.6 t         CH2       36.6 t       37.9 t         CH2       35.1 t       36.6 t         CH2       26.3 t       35.1 t         CH2       26.3 t       35.1 t         CH2       25.3 t       25.2 t         CH3       14.1 q       14.1 q	СН	72.7 d	72.6 d			
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$\begin{array}{c} CH_{3} \\ CH_{4} \\ CH_{4} \\ 114 \alpha \\ 114 $	CH <sub>2</sub>	17.9 a	17.9 a			
$CH_{-} \qquad 114a \qquad 114a$	CH <sub>2</sub>	14.1 a	14.1 a			
	CH,	11.4 a	11.4 a			

Table 1. <sup>13</sup>C NMR data of eurocidins D and E and their comparison with those of amphotericins A and B.

<sup>a</sup> In literature<sup>3)</sup>.

128 ~ 137 ppm region. The numbers of multiplicity of carbons were >C<; 4,  $-C_1^{LH}$ ; 24,  $>CH_2$ ; 9 and  $CH_3$ ; 3 for eurocidin D and >C<; 4,  $-C_1^{LH}$ ; 23,  $>CH_2$ ; 10 and  $CH_3$ ; 3 for eurocidin E, respectively. Consequently, it was found that one of methylenes of eurocidin E changed to a methine of eurocidin D, probably by the substitution of a hydrogen atom with a hydroxyl group.

HORII *et al.*<sup>2)</sup> reported that the partial structure of eurocidin A, which is purified from Eurocidin-T produced by *Streptomyces albireticuli*, contained 40 carbon atoms, pentaenic double bonds, a 30membered lactone ring, a secondary butyl side chain, a carboxyl group, a mycosamine moiety and some hydroxyl groups. The structures of eurocidins D and E were similar to that of eurocidin A, with regard to the total carbon number and two of four singlet carbon signals (<sup>13</sup>C NMR data). The two singlet carbon signals among four ones were defined as a lactone carbon and a carboxyl one. The comparison of the <sup>13</sup>C and <sup>1</sup>H NMR data of eurocidins D and E (Tables 1 and 2) with those of amphotericins A and B<sup>3,4)</sup>, heptaene macrolide antibiotics, suggests that the remaining two singlet carbon signals correspond to those of a ketone and a hemiketal.

## <sup>1</sup>H NMR and <sup>1</sup>H-COSY Spectra

The assignment of the protons of eurocidins D and E molecules is shown in Table 2. The values of the multiplicity were obtained by reffering to <sup>1</sup>H J resolved 2D NMR spectra, when clear multiplicity was observed, such as 32-CH<sub>3</sub>, 33-CH<sub>3</sub> and 6'-CH<sub>3</sub> *etc.* The <sup>1</sup>H-<sup>1</sup>H connectivity map (<sup>1</sup>H-COSY) for eurocidin E is shown in Fig. 2, marked with the connectivities for the regions of C-6 to C-10 and C-28 to C-33. Without references to further structural information the coupling sequences for the following parts of the molecule were able to be correctly interpreted: C-2 to C-4, C-6 to C-10, C-12 to C-18, C-29 to C-33 and mycosamine moiety. The cross peaks and chemical shifts of the regions of C-12 to C-18 and the mycosamine (C-1' to C-6') almost agreed with the data of amphotericins A and B<sup>3</sup>. Therefore it was estimated that the region of C-11 to C-18 contained a hemiketal (C-11) and C-14 (with a carboxyl group) had the same partial structure as amphotericins A and B, as well as mycosamine binding site at C-17.

Some polyenes such as nystatin  $A_1$ , pimaricin, tetrins A and  $B^{5 \sim 7}$ , as well as amphotericins A and B had the same partial structure as the region of C-11 to C-18 of eurocidin E. The common structures of such polyenes are suggested to be a mycosamine moiety, a hemiketal and a carboxyl group. The <sup>1</sup>H-COSY spectra of the region of C-2 to C-10 revealed that the cross peak connectivities were separated into two parts of the regions of C-2 to C-4 and C-6 to C-10. C-3 is considered to be a ketone carbon reffered to both chemical shifts of 2-H and 4-H protons and <sup>13</sup>C NMR data. Although it is possible that the carbons of C-2 to C-10 are sequenced in the reverse direction, the chemical shifts of 2-H (2.78 ppm) and 10-H protons (1.48 ppm) suggests that C-2 and C-10 are adjacent to the lactone and hemiketal carbons, respectively.

The comparison between eurocidins D and E in their <sup>1</sup>H-COSY spectra revealed that the cross assignments are identical with each other except for the region of C-6 to C-10. It is considered that one methylene-proton among C-6 to C-10 in eurocidin E had changes to a hydroxyl group in eurocidin D. <sup>1</sup>H NMR partial spectra  $(1.2 \sim 3.0 \text{ ppm})$  of eurocidins D and E are shown in Fig. 3. A doublet signal of two 8-H protons at 1.13 ppm in eurocidin E disappeared, and a doublet 8-H proton signal appeared at 2.98 ppm in eurocidin D (Fig. 3B and Table 2). The chemical shifts of methylene protons of 6-H, 7-H and 10-H of eurocidin D were not equivalent, and a 9-H proton signal was changed from broad singlet at 3.90 ppm in eurocidin E to a doublet at 3.80 ppm in eurocidin D. Therefore, the structure of eurocidin D

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Proton <sup>b</sup>	Chemical shift (ppm), multiplicity $(J(Hz))^a$				
	Eurocidin D	Eurocidin E	Amphotericin A <sup>c</sup>	Amphotericin B	
2-H	2.26	2.28 m	2.27	2.12	
2-H	2.47	2.45 m	2.34		
3-H	4.17	4.16 m	4.03	4.07	
4-H	2.26	2.28 m			
4-H	2.47	2.45 m			
6-H <sub>2</sub>		1.60			
6-H	2.39				
6-H	2.57				
7-H	1.28	2.26			
7-H	1.66	2.42			
8-H <sub>2</sub>		1.13 d (5.4)	1.50		
8-H	2.98 d (11.7)	. ,			
9-H	3.80 d (11.6)	3.90 br s	4.22	4.22	
10-H <sub>2</sub>		1.48 d (15.0)	1.60		
10-H	1.38 d (16.6)				
10-H	1.79				
12-H	1.10	1.10	1.13	1.10	
12-H	1.82	1.81	1.88	1.80	
13-H	3.99 m	3.98 m	3.95	3.96	
14-H	1.87	1.87 dd (10.4, 10.3)	1.88	1.86	
15-H	4.17	4.16 m	3.95	4.17	
16-H	1.51	1.53	1.75	1.47	
16-H	2.12	2.14	1.77	2.18	
17-H	4.38 br s	4.39 br s	4.34	4.28	
18-H	5.89 dd (15.6, 8.5)	5.87 dd (15.1, 8.4)	5.75	5.94	
19-H~26-H	6.00~6.40	6.00~6.40			
27-H	5.61 m	5.60 m	5.69		
28-H	2.22	2.20 m			
	2.32	2.34			
29-H	4.79 dd (10.7, 4.1)	4.80 dd (10.2, 3.0)			
30-H	1.54	1.57 m			
31-H	1.10	1.10			
	1.35 ddd (13.2, 7.9,	1.35 ddd (11.2, 7.7,			
	5.2)	4.8)			
32-CH <sub>3</sub>	0.85 t (7.3)	0.84 t (7.4)			
33-CH <sub>3</sub>	0.86 d (6.8)	0.85 d (6.8)			
Mycosamine		• •			
1'-H	4.47 s	4.51 s	4.47	4.46	
2'-H	3.70 d (1.5)	3.72 d (2.8)	3.70	3.79	
3'-H	2.74 d (8.8)	2.77 d (7.5)	2.68	2.96	
4'-H	3.12	3.14	3.07	3.18	
5'-H	3.20 m	3.23 m	3.14	3.24	
6'-CH-	1 17 d (6.0)	1.17 d (6.1)	1.13	1.15	

Table 2. <sup>1</sup>H NMR data of eurocidins D and E and their comparison with amphotericins A and B.

<sup>a</sup> When clear multiplets were observed, they are indicated here.

<sup>b</sup> Two numbers at one carbon indicate that the two hydrogens on the same carbon are not equivalent.

° In literature<sup>3)</sup>.

is proposed that which a hydroxyl group of 8-position is substituted for a 8-H proton of eurocidin E as shown in Fig. 1.

Both eurocidins D and E are proposed to have novel pentaenic structures revealed at the first time among eurocidin family, and eurocidin D is proposed as a novel substance. On the other hand it was not found that eurocidin E was different from eurocidin A, a component of Eurocidin-T, from several



Fig. 2. Contour plot of a 2D correlated <sup>1</sup>H NMR spectrum and <sup>1</sup>H-<sup>1</sup>H correlation map of eurocidin E (C-6 to C-10 and C-29 to C-33 regions).

HPLC analyses in the preceding paper<sup>1)</sup> and some structural data<sup>2)</sup>, but the producing microorganisms of these two compounds and their productive patterns of eurocidin family were different from each other. Eurocidin E may be an isomer of eurocidin A such as a position isomer of hydroxyl groups or a stereoisomer. Whether eurocidin E is identical to eurocidin A remains still unknown until elucidating the structure and the properties of purified eurocidin A from Eurocidin-T.

The configuration of asymmetric carbon atoms of eurocidins D and E is still unknown. The biological effects of eurocidins C, D and E on rat peritoneal mast cells compared with other polyenes, as well as their anti-microbial activities will be reported elsewhere<sup>8)</sup>.

#### Experimental

#### Chemicals

Reference substance was a gift from Takeda Chemical Industries Ltd., Japan, designated as Eurocidin-T in this paper. DMSO- $d_6$  (99.95% purity) was purchased from E. Merck, Darmstadt, FRG.

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NMR Spectrometry

NMR spectra were obtained on a Jeol JNM GX-400 spectrometer operating at 400 MHz with <sup>1</sup>H NMR and <sup>1</sup>H-<sup>1</sup>H shift correlated 2D NMR (<sup>1</sup>H-COSY). <sup>13</sup>C NMR, DEPT experiments ( $\theta$ =45°, 90° and 135°) and <sup>1</sup>H J resolved 2D NMR were recorded with Jeol JNM GX-270 spectrometer operating at 270 MHz. The amounts of sample used were about 10 mg in 1.0 ml of DMSO-d<sub>6</sub> for <sup>1</sup>H experiments and about 25 mg in 0.7 ml of DMSO-d<sub>6</sub> for <sup>13</sup>C experiments. Chemical shifts were given in ppm using DMSO-d<sub>6</sub> as the internal standard.

<sup>1</sup>H-COSY spectra were measured by the use of a 2D correlation sequence with a 90° mixing pulse. Data processing was carried out with the standard Jeol soft ware. An f2 spectral width of 2,500 Hz over 1,024 data points gave a digital resolution of 4.88 Hz. A total of 512 spectra, each of 32 transients, gave, with appropriate incrementing of the evolution delay, an f1 width of 2,500 Hz and a digital resolution of 4.9 Hz (with zero filling). The <sup>1</sup>H J resolved 2D spectra were obtained with the usual pulse sequence. The spectral widths were 1,170.1 Hz in f2 and 50.0 Hz in f1, giving a digital resolution of 1.14 and 0.1 Hz, respectively, and the data points were 2,048 × 512 matrix each of 64 transients.

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